**Lycopene6: Miniprep**

**source**: mini shaker

**samples**

*culture label side\_label destination plasmid*

K1-A pLYC33K-A K1-A Box\_Lyc6/A7 pLYC33K

K1-B pLYC33K-B K1-B Box\_Lyc6/B7 pLYC33K

**Protocol**

* Take an image of the culture block, upload it later to the Github issue
* For each sample, perform 2 minipreps (each on 2mL culture)
* Combine the two identical eluted minipreps in a regular zymo cleanup;

elute with **20uL** ddH2O

**Lycopene6: Sequence**

**source**:

sLYC10 benchtop/lyophilized

**samples**

*label location plasmid oligo*

K1-A Box\_Lyc6/A7 pLYC33K-A sLYC10

K1-B Box\_Lyc6/B7 pLYC33K-B sLYC10

**dilutions**

*label concentration destination*

sLYC10 100uM TPcon4\_stocks/G8

2.66uM sLYC10 2.66uM Box\_Lyc6/F5

**protocol**

* Make 100uM stock of sLYC10
* Make a 2.66uM stock of sLYC10:
  + 97.3 uL ddH2O
  + 2.7 uL oligo
* For each plasmid listed, mix the following sequencing reactions in an eppendorf tube:
  + 4 uL ddH2O
  + 6 uL miniprep DNA (undiluted)
  + 3 uL oligo (2.66 uM)
* Clearly label the tops of the tubes with the “label”
* Take the sequencing reactions and order form to:
  + 237 Stanley Hall (second floor cold room)